

Zooplankton and Microplankton Data
collected in the Gulf of Maine,
R.V. EASTWARD, June, 1977 and
R.V. CHALLENGE, July, 1979

edited by

V. Jones and F. Setchell

Bigelow Laboratory for Ocean Sciences

Technical Report No. 4

TABLE OF CONTENTS

	<u>Page</u>
I. Participants	1.
II. Methods	2.
A. R.V. EASTWARD	2.
1. Phytoplankton ETS	
2. Phytoplankton NR	
3. Zooplankton ETS	
B. R.V. CHALLENGE	4.
1. Phytoplankton ETS	
2. Phytoplankton protein	
3. Phytoplankton chlorophyll	
4. Experiment on ETS method precision	
III. References	6.
IV. R.V. EASTWARD cruise track (Fig. 1)	8.
V. Microplankton data (Table E-1)	10.
VI. Integrated and averaged microplankton data (Table E-2)	11.
VII. Respiratory O ₂ Consumed and CO ₂ Evolved (Table E-3)	12.
VIII. Fractionation data (Table E-4)	13.
IX. Zooplankton data (Table E-5)	14.
X. R.V. CHALLENGE station locations (Fig. 2)	15.
XI. Microplankton ETS, chlorophyll, and protein (Table C-1)	16.
XII. Regression of ETS vs. protein (Fig. 3)	18.
XIII. Respiratory O ₂ Consumed and CO ₂ Evolved (Table C-2)	19.
XIV. Integrated microplankton ETS and chlorophyll (Table C-3)	20.

PARTICIPANTS

R/V EASTWARD

F.D. KING

Zooplankton ETS,
Phytoplankton ETS

C. SAPIENZA

Phytoplankton ETS,
NR*R/V CHALLENGE*

T.T. PACKARD

Phytoplankton ETS

F.W. SETCHELL

Phytoplankton ETS,
protein

Chlorophyll-*a* data was generously provided by I. Morris for the *R/V EASTWARD* cruise and D. Phinney for the *R/V CHALLENGE* cruise.

METHODS

R/V EASTWARDPhytoplankton

ETS: Four liter samples were drawn from Niskin bottles, filtered onto 47mm Gelman A/E glass fiber filters, placed in numbered vials, and frozen in liquid nitrogen (Ahmed *et al.*, 1976). The samples were assayed using a semi-automated procedure (Abrahamson *et al.*, 1980) designed from the technique of Kenner and Ahmed (1975a) after return to the laboratory. ETS activity was corrected to *in situ* temperature using the Arrhenius equation and an activation energy of 15 Kcal. mole (Owens and King, 1975).

NR: Four liter samples were drawn from Niskin bottles and assayed immediately following the method of Eppley, Coatsworth, and Solorzano (1969). A standard curve for NO_2^- concentration was determined after the method described by Bass and Packard (1977).

Chlorophyll: was measured fluorometrically following the technique of Holm- Hansen *et al.*, (1965).

Calculations

- 1) Integrated ETS, NR, and chlorophyll were determined by trapezoidal approximation.
- 2) Average concentrations or activity was found by dividing the integrated values by the interval of integration in meters.

- 3) O_2 consumption was calculated from ETS values using the R/ETS factor of 0.15 (Kenner and Ahmed, 1975b).
- 4) O_2 consumption was converted to CO_2 evolution using an R/Q of 1 and the mass conversion factor of 0.536 (12/22.4).

Zooplankton

ETS: was measured using the method of Owens and King (1975) on the automated system described by Abrahamson *et al.* (1980). The organisms were collected with a 212 μ M mesh net, 1m in diameter. A Folsom plankton splitter was used to divide the collected material. One quarter of the haul was placed in a numbered plastic vial, promptly frozen in liquid nitrogen, and returned to the laboratory for ETS analysis. ETS activity was corrected to average *in situ* temperature for the depth interval sampled using the Arrhenius equation and an activation energy of 15 Kcal·mole. (Owens and King, 1975).

Calculations

- 1) Zooplankton O_2 consumption was calculated using the R/ETS factor of Owens and King (1975).
- 2) CO_2 evolution was calculated using an R/Q of 0.85 and the mass conversion of 0.536 (12/22.4).
- 3) NH_4^+ excretion was calculated from O_2 consumption using an O:N atomic ratio of 8:1 (Smith and Whitledge, 1977 and Packard, 1979).

METHODS

R/V CHALLENGE

ETS: was measured on 2-4 liter samples from a submersible pumping system (Holligan and Harbor, 1977) or from discrete samples after the method of Kenner and Ahmed (1975a) using the quench solution suggested by Owens and King (1975). All assay were performed on board.

PROTEIN: a portion of the cleared ETS homogenate was frozen and returned to the laboratory where protein was measured using Setchell's (submitted manuscript) modification of Bradford's (1976) coomassie blue dye binding assay.

CHLOROPHYLL: was measured fluorometrically after the method of Yentsch and Menzel (1963) with no correction for phaeopigments.

Calculations:

- 1) Respiration was calculated from ETS data in the same manner as was done with the *R/V EASTWARD* data.

Experiment to test precision of ETS method: (T. Packard)

Four liters of surface water were collected off Squirrel Island and filtered through a Gelman glass fiber filter. A 5.5 μ l homogenate was prepared and four (4) replicates were run through the Kenner and Ahmed (1975) assay on the centrifuged homogenate. The absorbances in a 1 cm path-length cell were 0.30, 0.31, 0.30, and 0.30 as measured by the minispec. The average is 0.30 and the standard deviation is 0.01. This is a 3.3% error.

- Abrahamson, J., F. Setchell, V. Jones, and T.T. Packard (1980) Automated chemical analysis for measuring respiratory electron transport activity in marine plankton. CUEA Tech. Rept. 48.
- Ahmed, S.I., R.A. Kenner, and F.D. King (1976) Preservation of enzymic activity in marine plankton by low-temperature freezing. Mar. Chem. 4: 133-139.
- Bass, A.E. and T.T. Packard (1977) Physical, chemical, and biological observations from JOINT-II R.V. Alpha Helix Leg 0, 5-20 March 1976. CUEA Data Rept. 41.
- Bradford, M.M. (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein using the principle of protein dye binding. Anal. Biochem. 72: 248-254.
- Eppley, R.W., J.L. Coatsworth, and L. Solorzano (1969) Studies of nitrate reductase in marine phytoplankton. Limnol. Oceanogr. 14: 194-205.
- Holligan, P.M. and D.S. Harbor (1977) The vertical distribution and succession of phytoplankton in the western English Channel in 1975 and 1976. J. Mar. Biol. Assn. U.K. 57: 1075-1093.
- Holm-Hansen, O., C.J. Lorenzen, R.W. Holmes, and J.D.H. Strickland (1965) Fluorometric determination of chlorophyll. J. du Conseil Conseil perm. internatl. pour l'explor. de la mer 30: 3-15.
- Kenner, R.A. and S.I. Ahmed (1975a) Measurement of electron transport activities in marine phytoplankton. Mar. Biol. 33: 119-127.
- Kenner, R.A. and S.I. Ahmed (1975b) Correlation between oxygen utilization and electron transport activity in marine phytoplankton. Mar. Biol. 33: 129-133.

- Owens, T.G. and F.D. King (1975) The measurement of respiratory electron transport system activity in marine zooplankton. Mar. Biol. 30: 27-36.
- Packard, T.T. (1979) Respiration and respiratory electron transport activity in plankton from the Northwest African upwelling area. CUEA Tech. Rept. 47.
- Setchell, F. (submitted) Particulate protein in coastal waters of the Gulf of Maine. To J. Mar. Chem.
- Smith, S.L. and T.E. Whitley (1977) The role of zooplankton in the regeneration of nitrogen in a coastal upwelling system off northwest Africa. Deep Sea Res. 24: 49-56.
- Yentsch, C.S. and D.W. Menzel (1963) A method for the determination of phytoplankton chlorophyll and phaeophytin by fluorescence. Deep Sea Res. 10: 221-231.

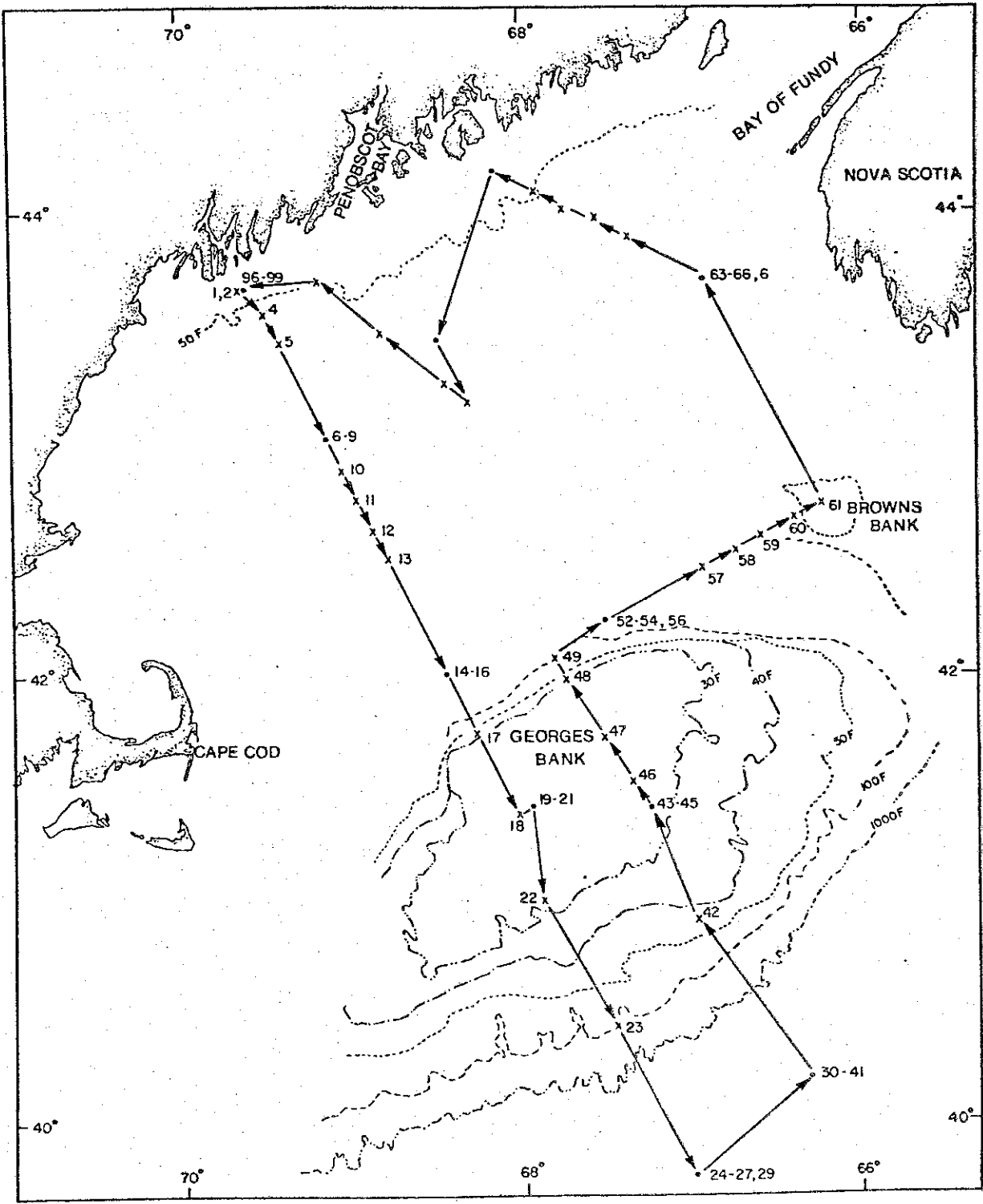


Fig. 1. Cruise track, R.V. EASTWARD, June, 1977.

Table E-1. Summary of microplankton ETS, NR and Chlorophyll a data by depth. R/V EASTWARD
cruise during June-July 1977.

DATE	STN. NO.	POSITION	DEPTH m	%I.	TEMP. IN SITU	INCUBATION	NR ACTIVITY ($\mu\text{M NO}_2 \times \text{h}^{-1}$)	ETS ACTIVITY ($\mu\text{g O}_2 \times \text{h}^{-1}$)	CHL- a ($\mu\text{g} \times \text{h}^{-1}$)	NR/ CHL	ETS/ CHL
6/28	8	56°28.00N 156°00.00W	0	100	9.3	13.9	9.59	0.84	3.92	2.45	0.21
			6	40	9.5	↓	6.01	1.17	3.98	1.51	0.29
			6	20	7.0	↓	19.41	0.61	4.45	4.36	0.14
			0	100	14.4	↓	n.d.	2.03	0.38	n.d.	5.34
			3	40	14.4	↓	n.d.	2.25	0.09	n.d.	25.00
			6	20	14.3	↓	n.d.	2.56	0.37	n.d.	6.92
6/29	15	00°00.00N 156°27.00W	0	100	14.4	↓	n.d.	2.03	0.38	n.d.	5.34
			3	40	14.4	↓	n.d.	2.25	0.09	n.d.	25.00
			6	20	14.3	↓	n.d.	2.56	0.37	n.d.	6.92
			9	10	14.2	↓	n.d.	1.64	0.42	n.d.	3.90
			13	3	13.1	↓	n.d.	2.08	0.28	n.d.	7.43
			17	1	11.4	↓	n.d.	1.51	0.73	n.d.	2.07
6/30	20	25°00.00N 156°56.50W	0	100	12.3	14.4	0.50	4.76	3.12	0.16	1.53
			3	40	12.3	↓	0.39	4.29	2.17	0.18	1.98
			5	20	12.3	↓	0.57	4.44	2.17	0.26	2.05
			7	10	12.4	↓	0.47	5.24	2.23	0.21	2.35
			10	4	12.4	↓	0.87	4.00	2.44	0.36	1.64
			14	1	12.3	↓	0.13	6.77	2.33	0.06	2.91
7/1	26	45°00.00N 158°58.90W	0	100	20.0	20.0	0.12	2.19	0.13	0.92	16.85
			4	40	19.9	↓	0.50	3.35	0.23	2.17	14.56
			8	20	19.8	↓	0.15	3.80	0.28	0.53	13.57
			21	10	18.8	↓	0.24	2.13	0.66	0.37	3.23
			33	4	17.2	↓	0.25	3.14	0.12	2.08	26.16
			51	1	14.7	↓	0.06	0.94	0.10	0.62	9.40
7/2	33	13°00.00N 155°15.50W	5	40	n.d.	21.5	0.07	2.01	0.14	0.50	14.36
			35	3	n.d.	↓	0.32	0.65	0.80	0.40	0.81
			45	1	n.d.	↓	0.85	6.99	1.80	0.47	3.88
			0	100	11.5	11.5	0.78	3.20	2.33	0.33	1.37
			3	40	11.5	↓	0.35	3.52	1.80	0.19	1.96
			6	20	11.3	↓	0.38	4.47	2.39	0.16	1.87
7/3	45	25°25.50N 155°15.50W	8	10	11.0	11.5	0.59	3.54	3.29	0.18	1.08
			12	4	10.7	↓	0.41	4.74	2.76	0.15	1.72
			17	1	10.7	↓	0.37	4.92	2.54	0.15	1.94

STN. NO.	POSITION	DEPTH		TEMP. °C	INCUBATION	NR ACTIVITY (nm NO ₂ ¹h ⁻¹)	ETS ACTIVITY (μg O ₂ ¹h ⁻¹)	CHL-a (μg ¹h ⁻¹)	NR/CHL	ETS/CHL	
		m	% I								
4	54	↕	0	100	14.5	14.5	0.33	2.12	0.38	0.87	5.58
			4	40	14.2		0.26	2.32	0.52	0.50	4.46
			9	20	13.7		0.20	3.72	0.52	0.38	7.15
			14	10	12.8		0.43	3.09	0.74	0.58	4.17
			21	4	11.4		0.71	2.46	1.12	0.64	2.19
			31	1	9.5	0.47	3.54	0.78	0.60	4.54	
5	63	↕	0	100	13.3	13.5	0.13	4.03	0.88	0.15	4.58
			4.5	40	11.7		0.14	3.96	0.94	0.15	4.21
			8	20	10.3		0.17	3.37	1.14	0.15	2.96
			12	10	9.3		0.20	2.63	1.14	0.18	2.30
			16	4	9.1		0.42	3.42	1.00	0.42	3.42
			25	1	8.2	0.25	1.62	0.90	0.28	1.80	
6	76	↕	0	100	9.5	16.8	1.47	2.22	2.33	0.63	0.95
			2.5	40	9.0		1.58	2.77	2.44	0.65	0.14
			4.5	20	8.6		1.04	2.55	2.97	0.35	0.86
			7	10	8.2		1.95	3.24	2.86	0.68	1.13
			10	4	8.1		1.03	3.19	3.02	0.34	1.06
			17	1	7.3	0.51	1.06	1.59	0.32	0.67	
7	86	↕	0	100	13.1	14.8	0.09	1.65	0.68	0.13	2.42
			5	40	12.6		0.16	1.62	0.64	0.24	2.54
			10	20	12.1		0.56	2.43	0.88	0.64	2.76
			14	10	11.6		0.38	2.12	0.98	0.39	2.16
			20	4	11.0		1.11	1.80	1.32	0.84	1.37
			29	1	7.3	0.47	1.28	1.04	0.45	1.23	
8	97	↕	0	100	12.9	12.5	0.33	3.25	0.64	0.52	5.08
			3	40	12.5		0.29	2.07	0.84	0.35	2.47
			5	20	12.2		0.50	0.39	0.84	0.59	0.46
			8.5	10	11.6		0.51	2.58	1.00	0.51	2.58
			13	4	10.9		1.65	1.73	1.42	1.17	1.22
			21	1	9.3	0.46	3.05	1.34	0.34	2.28	

Table E-2. Areal and mean chlorophyll, ETS, AND NR. R, V EASTWARD, June, 1977. Areal measurements are for the euphotic zone, 100% to 1% I₀, except as noted. Mean values are areal values divided by depth interval. Units are Chlorophyll-a = mg m⁻²; mean Chlorophyll-a = μg l⁻¹; ETS = m% O₂ m⁻² h⁻¹; mean ETS = μ% O₂ l⁻¹ h⁻¹; ETS/Chl = μ% O₂ · μg Chl⁻¹ h⁻¹; NR = μM NO₂ l⁻¹ h⁻¹; mean NR = nM NO₂ l⁻¹ h⁻¹; NR/Chl = nM NO₂ · μg Chl. These values are calculated from the data presented in Table E-1.

STN. NO.	MAX (m)	ΣCHL-α	MEAN CHL	ΣETS	MEAN ETS	ΣETS / ΣCHL	ENR	MEAN NR	ENR / ΣCHL-α
8*	6	24.36	4.06	5.74	0.96	0.24	59.08	9.85	2.43
15	17	6.00	0.35	34.56	2.03	5.76	--	--	--
20	14	33.22	2.37	67.39	4.81	2.03	7.35	0.52	0.22
26	51	14.51	0.28	132.27	2.59	9.12	10.81	0.21	0.75
33†	45	27.80	0.62	88.15	1.96	3.17	12.05	0.27	0.43
45	17	43.51	2.56	70.79	4.69	1.63	7.71	0.45	0.18
54	31	23.55	0.76	90.43	2.92	3.84	13.80	0.45	0.59
63	25	25.13	1.01	77.59	3.10	3.09	6.15	0.25	0.24
76	17	43.62	2.57	43.32	2.55	0.99	20.03	1.18	0.46
86	29	28.34	0.98	53.02	1.83	1.87	15.89	0.55	0.56
97	21	23.61	1.12	44.46	2.12	1.88	16.79	0.80	0.71

*Samples from 100% to 20% light depth only (0m, 3.5m, and 6m)

†Samples from 40%, 3% and 1% light depth only (5m, 35m and 45m). This number includes a 0-5m average based on the value at the 40% light depth.

Table E-3. Respiration in terms of O₂ consumed and CO₂ evolved. Values are calculated on an areal basis and on a mean euphotic zone basis. Integrated and average ETS values in Table E-2 were used for these calculations.

STN NO.	O ₂ CONSUMED (ml O ₂ m ⁻² h ⁻¹)	AVERAGE O ₂ CONSUMED (μl O ₂ l ⁻¹ h ⁻¹)	CO ₂ EVOLVED (mg C · m ⁻² h ⁻¹)	AVERAGE CO ₂ EVOLVED (μg C · l ⁻¹ h ⁻¹)
8*	0.86	0.14	0.46	0.08
15	5.18	0.30	2.78	0.16
20	10.11	0.72	5.42	0.39
26	19.84	0.39	10.63	0.21
33+	13.22	0.29	7.08	0.16
45	10.62	0.70	5.69	0.38
54	13.56	0.44	7.26	0.24
63	11.64	0.47	6.23	0.25
76	6.50	0.38	3.48	0.20
86	7.95	0.27	4.26	0.14
97	6.67	0.32	3.57	0.17

* Samples from 100%, 40% and 20% light depths only (0m, 3.5m and 6m)

+ Samples from 40%, 3% and 1% light depths only (5m, 35m and 45m). This number includes a 0-5m average based on the value at the 40% light depth.

Table E-4. Microplankton ETS, NR, and chlorophyll fractionation data done on samples taken from the 40% light depth during the R.V. EASTWARD cruise to the Gulf of Maine, June, 1977. These numbers have not been checked as the pertinent data were missing from the original data sheets.

STN. NO.	DEPTH (m)	SIZE FRACTION (μ)	TEMP. <i>in situ</i>	inc. (nm NO ₂ h ⁻¹ ¹)	NR (h ⁻¹ ¹)	ETS (h ⁻¹ ¹)	CHLOROPHYLL (μ g ¹)	NR/CHL	ETS/CHL
8	3.5	<10	9.5	13.9	--	1.67	0.86	--	1.94
15	3	<10	14.4	--	--	2.90	0.02	--	145.00
20	3	<10	12.3	14.4	0.17	3.21	0.42	0.41	7.64
26	4	<10	19.9	20.0	0.10	4.92	0.13	0.76	37.85
33	5	<10	--	21.5	0.04	4.13	0.02	2.00	206.50
45	3	<10	11.5	11.5	0.22	3.46	0.31	0.71	11.17
54	4	<10	14.2	14.5	0.15	3.08	0.26	0.56	11.83
63	4.5	<10	11.7	13.5	0.04	3.91	0.47	0.09	8.32
76	2.5	<10	9.0	16.8	0.05	0.81	1.46	0.03	0.55
86	5	<10	12.6	14.8	0.03	2.14	0.12	0.27	17.83
97	3	<10	12.5	12.5	0.28	2.34	0.46	0.61	5.09

Table E-5. Zooplankton ETS activity, O₂ consumption, CO₂ evolution, NH₄⁺ excretion, and dry weight of collected material. All hauls were done with a 212 μm mesh, 1 m diameter net. ETS activity was corrected to average in situ activity for the haul (Owens and King, 1975).

STATION NO.	DEPTH INTERVAL (m)	ETS (ml O ₂ m ⁻² h ⁻¹)	O ₂ CONSUMED (m ³ m ⁻² h ⁻¹)	CO ₂ EVOLVED (mg C m ⁻² h ⁻¹)	NH ₄ ⁺ (μg-at m ⁻² h ⁻¹)	DRY WEIGHT (mg m ⁻²)
8	30-0	5.76	2.88	1.31	32.14	24.7
8	150-30	25.44	12.72	5.79	141.96	47.8
8	150-30	27.72	13.86	6.31	154.69	47.8
19	30-0	23.61	11.81	5.38	131.81	117.2
26	30-0	0.66	0.33	0.15	3.68	4.3
26	100-30	0.32	0.16	0.07	1.80	0.62
26	200-100	0.41	0.21	0.09	2.29	0.054
30	30-0	1.47	0.74	0.33	8.26	5.84
45	30-0	18.66	9.33	4.25	104.13	--
45	50-30	11.66	5.83	2.65	65.07	78.6
54	30-0	70.23	35.12	15.98	391.91	122.3
54	100-30	41.86	20.93	9.53	233.59	75.2
54	250-100	15.90	7.95	3.62	88.72	15.8
63	30-0	19.77	9.89	4.50	110.32	65.9
63	130-30	94.60	47.30	21.54	527.90	41.1
76	30-0	0.96	0.48	0.22	5.36	22.0
76	60-30	7.02	3.51	1.60	39.17	2.5
85	30-0	89.94	44.97	17.84	501.90	171.2
85	155-30	46.13	23.06	10.50	257.39	38.6
97	30-0	46.53	23.27	10.59	259.65	42.1
97	70-30	0.80	0.40	0.18	4.46	13.8

July 17, 1979
 July 18, 1979
 July 19, 1979
 July 23, 1979
 July 25, 1979
 July 26, 1979

1-5
 6,7
 8,9
 10,11
 12,13
 14,15

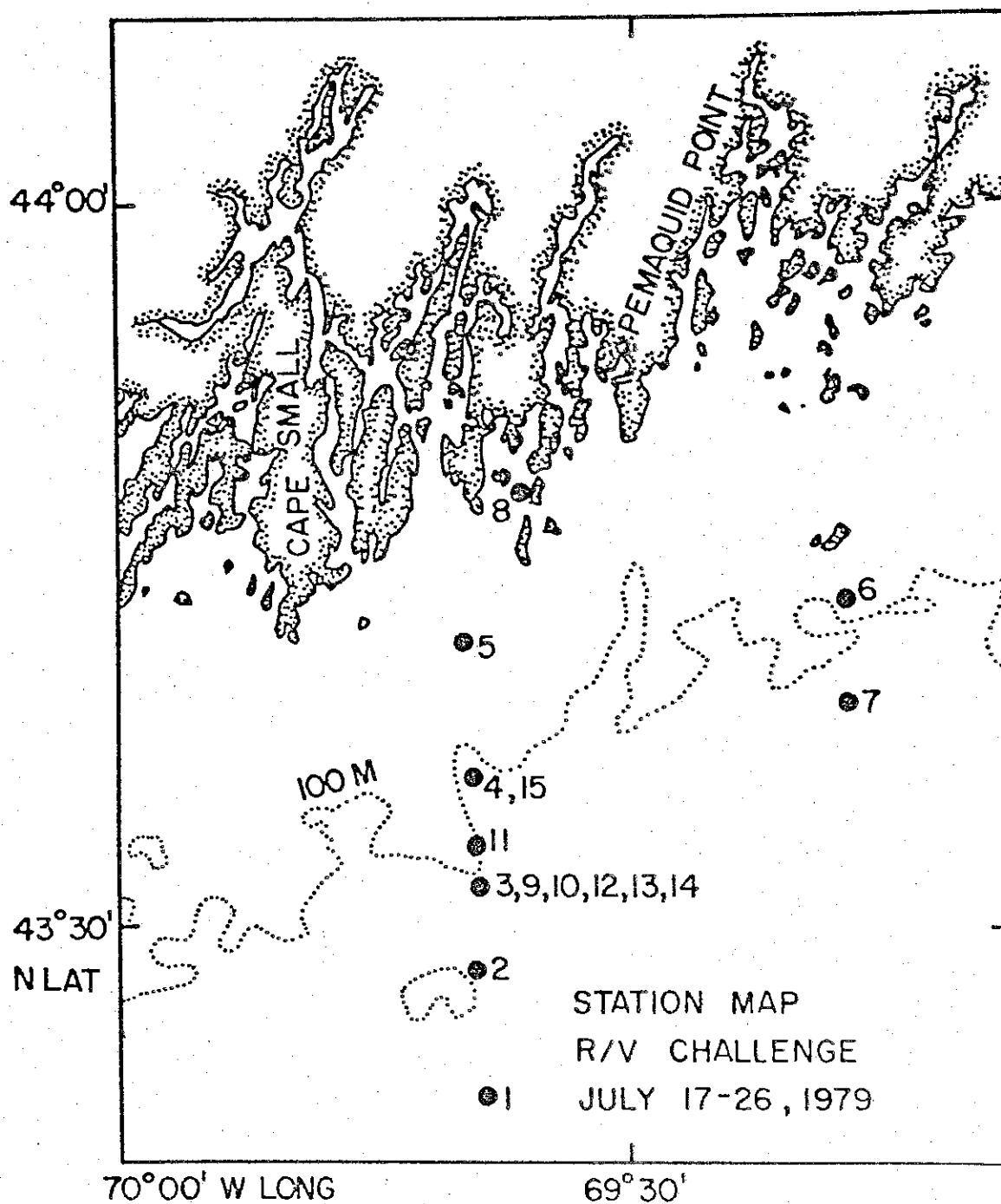


Fig. 2. Station locations, R.V. CHALLENGE cruise, July, 1979.

Table C-1. Summary of microplankton ETS, chlorophyll, and protein measurements for the R.V. CHALLENGE cruise, July, 1979. Protein N is calculated from protein by dividing by 87.5 (6.25 x 14).

STATION & SAMPLE NO.	DEPTH (m)	ETS ($\mu\text{g O}_2 \text{ l}^{-1}\text{h}^{-1}$)	TOTAL CHL ($\mu\text{g} \cdot \text{l}^{-1}$)	PROTEIN ($\mu\text{g} \cdot \text{l}^{-1}$)	PROTEIN N ($\mu\text{g at} \cdot \text{l}^{-1}$)	ETS/CHL	ETS/PROTEIN	ETS/PROTEIN N	N/CHL
8-1	34-20	1.31	1.43	24.43	0.28	0.92	0.05	4.68	0.20
2	20-2	2.46	3.26	110.32	1.25	0.75	0.02	0.97	0.38
9-1	60-49	-----	-----	2.02	0.02	-----	-----	-----	-----
2	49-23	2.65	-----	5.66	0.58	-----	0.47	4.57	-----
3	23-2	4.80	-----	77.13	0.88	-----	0.06	5.45	-----
4	60-49	-----	0.31	2.05	0.02	-----	-----	-----	.07
5	49-31	1.87	1.79	32.03	0.37	1.04	0.06	5.05	0.21
6	31-12	5.79	2.48	144.97	1.66	2.33	0.04	3.49	0.67
7	12-2	3.84	1.03	-----	-----	3.73	-----	-----	-----
10-1	74-45	0.70	0.50	2.02	0.02	1.40	0.35	35.00	0.04
2	45-25	*1.78	0.88	8.58	0.10	2.02	0.21	17.80	0.11
3	25-8	5.74	2.67	115.73	1.32	2.15	0.05	4.35	0.49
4	8-2	*5.31	0.99	61.41	2.70	5.36	0.09	1.97	2.72
5	36	0.34	1.04	4.86	0.06	0.33	0.07	5.67	0.06
6	29	1.15	1.34	10.73	0.12	0.86	0.11	9.58	0.09
7	18	9.94	4.59	212.22	2.43	2.17	0.05	4.09	0.53
12-1	62-54	*0.57	1.24	39.74	0.45	0.46	0.01	1.27	0.36
2	54-44	1.29	3.08	67.62	0.77	0.42	0.02	1.68	0.25
3	44-19	1.48	1.15	19.36	0.22	1.29	0.08	6.73	0.19
+	19-4	*1.94	2.37	64.37	0.74	0.82	0.03	2.62	0.31
+	5	*1.19	1.66	26.06	0.30	0.72	0.05	3.97	0.18
+	6	3.98	1.80	93.47	1.07	2.21	0.04	3.72	0.48
13-1	68-48	1.01	2.04	169.32	1.94	2.50	0.01	0.52	0.95
2	48-23	*0.61	1.38	16.28	0.19	0.46	0.04	3.21	0.14
3	23-6	2.81	1.89	76.69	0.88	1.49	0.04	3.19	0.47
4	6-2	*3.51	1.28	63.28	0.72	2.74	0.06	4.88	0.56

* Noted a few zooplankton in the sample. At Sta. 12-4, luminescence was noted as filter went dry.

† These samples were taken from Niskin bottles, samples 1-3 were taken from the submersible pumping system.

Table C-1 Con'td.

STATION & SAMPLE NO.	DEPTH (m)	ETS ($\mu\text{g O}_2 \text{ l}^{-1} \text{ h}^{-1}$)	TOTAL CHL ($\mu\text{g} \cdot \text{l}^{-1}$)	PROTEIN ($\mu\text{g} \cdot \text{l}^{-1}$)	PROTEIN N ($\mu\text{g at} \cdot \text{l}^{-1}$)	ETS / CHL	ETS / PROTEIN	ETS / N	N / CHL
14-1	70-52	1.14	0.36	19.98	0.22	3.17	0.06	5.18	0.61
2	52-29	*0.95	0.63	33.20	0.38	1.51	0.03	2.50	0.60
3	29-10	4.70	3.48	155.33	1.78	1.35	0.03	2.64	0.51
4	10-2	*3.03	1.09	52.71	0.60	2.78	0.06	5.05	0.55
15-1	64-26	1.19	1.04	31.37	0.36	1.14	0.04	3.31	0.35
Mean		2.66	1.66	58.07	.68	1.61	.08	6.94	38
		2.17	1.04	55.81	.63	1.20	.10	9.41	.25
		29	29	30	30	26	28	27	28

* A few zooplankton were noted in these samples.

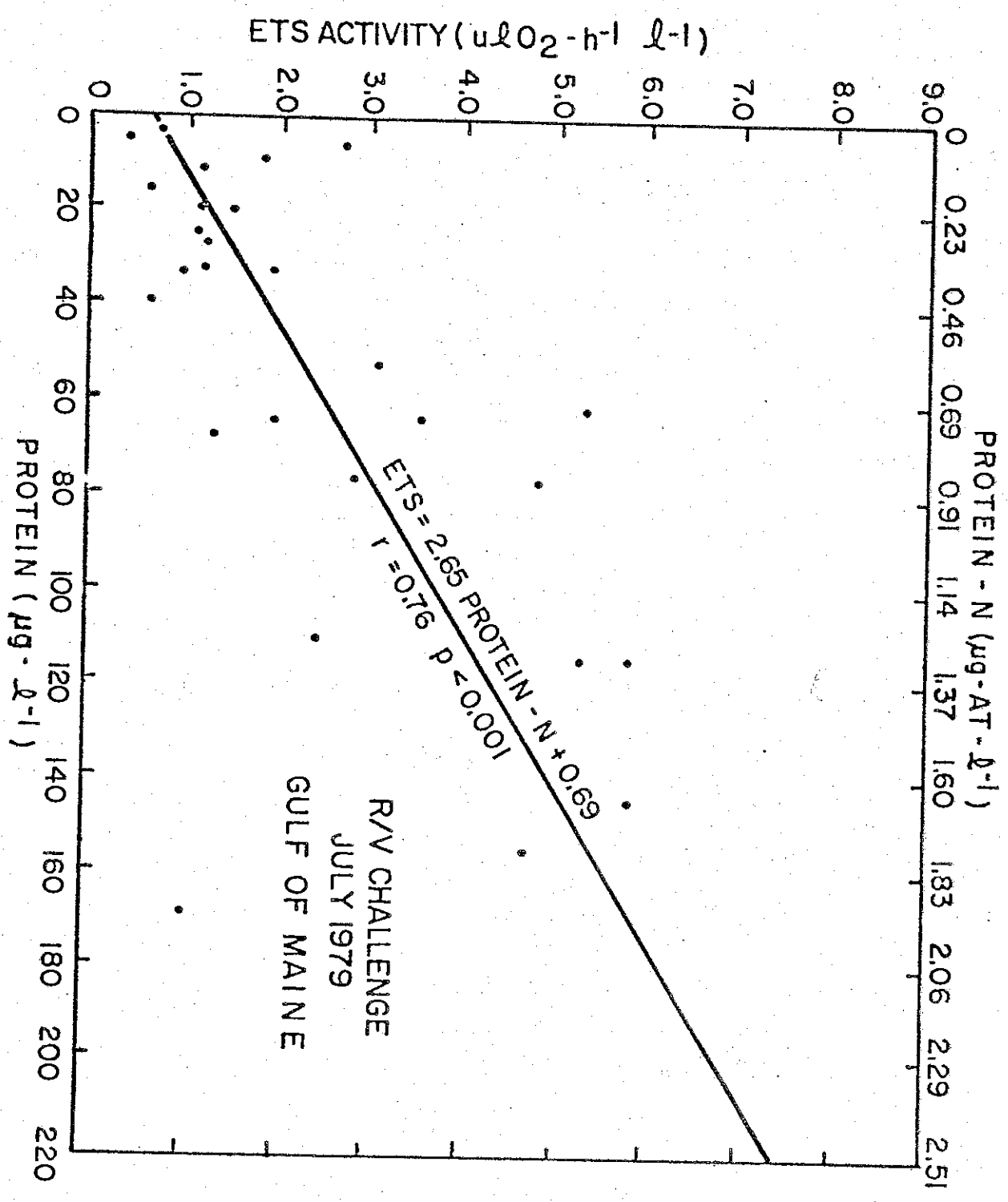


Fig. 3.

Table C-2. Respiratory O₂ Consumption and CO₂ Evolution from ETS data (Table C-1), R/V CHALLENGE, July 1979.

STATION AND SAMPLE NO.	DEPTH OR INTERVAL (m)	O ₂ CONSUMPTION (μl O ₂ l ⁻¹ h ⁻¹)	CO ₂ EVOLVED (μg C l ⁻¹ h ⁻¹)
8-1	34-20	0.20	0.11
2	20-2	0.37	0.20
9-1	60-49	--	--
2	49-23	0.40	0.21
3	23-2	0.72	0.39
5	49-31	0.28	0.15
6	31-12	0.87	0.47
7	12-2	0.58	0.31
10-1	74-45	0.11	0.06
2	45-28	0.27	0.14
3	28-8	0.86	0.46
4	8-2	0.80	0.43
5	36	0.05	0.03
6	29	0.17	0.09
7	18	1.49	0.80
12-1	62-54	0.09	0.05
2	54-44	0.19	0.10
3	44-19	0.22	0.12
4	19-4	0.29	0.16
5	29	0.18	0.10
6	14	0.60	0.32
13-1	68-48	0.15	0.08
2	48-23	0.09	0.05
3	23-6	0.42	0.23
4	6-2	0.53	0.28
14-1	75-52	0.17	0.09
2	52-29	0.14	0.08
3	29-10	0.71	0.38
4	10-2	0.45	0.24
15-1	69-26	0.18	0.10

Table C-3. Microplankton ETS activity in samples taken in the Gulf of Maine, July 17, 1979, during the R/V CHALLENGE cruise. Measurements were made by T. Packard using a mini-spectrophotometer, 1 cm cell, and 4 liters of filtrate. A blank consisted of 0.1 ml of filtered seawater ground with a glass fiber filter (A = 0.020). The "I" in the depth description columns means the values are integrated. Chlorophyll measurements were done by D. Pinney by the method described in Yentsch and Menzel (1963) with the exception that calibration is standardized against pure CHL-a (Sigma Scientific). Gary Grunseich has depth profiles.

STATION NO.	DEPTH DESCRIPTION	CORRECTED DEPTH INTERVAL	ETS ACTIVITY ($\mu\text{g O}_2 \text{ h}^{-1} \text{ l}^{-1}$)	CHLOROPHYLL ($\mu\text{g l}^{-1}$)	ETS/CHL	METHODS NOTES	BIOLOGY NOTES
1	Surface-I	14-2	1.81	0.75	2.41	Centrifugation done after incubation, temperature not exact	
	Chl-Max-I	33-14	---	2.18	---		
1	Bottom-I	52-33	1.46	0.88	1.67	Temp. = 10°C, Centrifugation done after incubation.	filter a light yellow color Small copepod unable to remove in sample
2	Surface-I	8.5-2	---	0.67	---		filter a golden brown color
2	Chl-Max-I	36-8.5	3.67	2.04	1.81	"	golden brown color
2	Chl-Max	16	15.65	2.56	6.11	"	
2	Bottom-I	56-36	1.95	0.53	3.68	Temp. not exact, centrifugation done after incubation	Removed 3 copepods
4	Surface -I	9-2	3.19	1.42	2.25	Centrifugation done before incubation; temp. exact	
4	Thermocline-II	26-9	4.67	3.04	1.54	"	Golden brown color
4	Thermocline-I	42-26	0.27	0.53	0.51	"	Filter went dry after 15 sec.
4	Bottom-I	60-42	0.29	0.38	0.76	"	